

RESUMEN

En esta Tesis doctoral se profundizó en el estudio de la interacción lípido-receptor de acetilcolina nicotínico (AChR) en dos aspectos: por un lado, el mecanismo de inhibición de ácidos grasos libres (AGLs), antagonistas no competitivos del AChR, y por el otro, la ubicación del AChR en dominios líquido-ordenados (L_o) condicionada por dos características particulares de la membrana.

Con la finalidad de dilucidar el mecanismo de antagonismo de los AGLs sobre el AChR, se utilizaron AGLs con un doble enlace único en diferentes posiciones de una cadena acílica de 18 átomos de carbono. Estudios funcionales realizados con la técnica de *patch-clamp* han mostrado que solo el *cis*-6-18:1 y el *cis*-9-18:1 reducen la duración del estado de canal abierto del receptor, sugiriendo, por lo tanto, un mecanismo de bloqueo alostérico del canal. Mediante espectroscopía de fluorescencia se comprobó que todos los AGLs se ubican en la interfase lípido-AChR, quedando el *cis*-6-18:1 restringido a los sitios denominados sitios anulares, mientras que el resto de los AGLs ocupa también sitios no-anulares. Por otro lado, estudios de polarización de fluorescencia mostraron que el AGLs *cis*-9-18:1 es el que ocasiona el mayor desorden en la membrana. Se comprobó i) que todos los *cis*-AGLs generan cambios conformacionales del AChR a nivel transmembrana, ii) que los *cis*-9-18:1, *cis*-11-18:1 y *cis*-13-18:1 perturban al AChR en su estado de reposo e iii) que los *cis*-6-18:1 y *cis*-9-18:1 son los que causan una mayor perturbación del estado desensibilizado. De esta manera, la posición e isomería del ángulo de torsión de los AGLs insaturados serían un factor clave en el bloqueo del AChR, sugiriendo entonces que los AGLs con un único doble enlace y ubicados superficialmente en la membrana inhiben en forma directa la función del AChR, posiblemente al perturbar una secuencia aminoacídica

transmembrana involucrada en los cambios alostéricos necesarios para la apertura del canal iónico.

Se postula que en la membrana plasmática el AChR se encuentra en dominios lipídicos denominados “balsas” (“*rafts*”). Sin embargo, el AChR no muestra preferencia por dominios L_o en sistemas modelo compuestos por esfingomielina (SM), colesterol (Col) y POPC (1:1:1), pero sí lo hace un segmento transmembrana (γ M4) que exhibe mayor contacto con los lípidos. Es decir, su distribución no dependería exclusivamente de sus propiedades intrínsecas sino también de señales extrínsecas a la proteína. En este trabajo de Tesis se estudió la posible partición diferencial del AChR en los dominios L_o en dos sistemas modelo diferentes en función de: a) la presencia de diferentes especies puras de SM en la membrana, y b) la existencia de asimetría transbica en el sistema modelo, mediante el agregado de SM de cerebro (bSM) en la hemicapa externa. Tanto la existencia de asimetría como la presencia de 16:0-SM o 18:0-SM, en comparación con las bSM y 24:1-SM, producen una partición preferencial del AChR en los dominios L_o . De este modo, la localización del AChR en estos dominios depende no solo de sus propiedades sino también de las características propias de la membrana en la que se encuentra.

Entender la interacción lípido-AChR es de gran importancia para determinar tratamientos que puedan mejorar o inhibir la función del AChR y tratar enfermedades que lo involucren.

ABSTRACT

In this Ph. D. thesis the understanding of the lipid-nicotinic acetylcholine receptor (AChR) interaction was furthered in two aspects, namely the inhibition mechanism of free fatty acids (FFA), non-competitive AChR antagonists, and AChR location in liquid-ordered (L_o) domains conditioned by two membrane characteristics.

To elucidate FFA's antagonism mechanism, FFA with only one double bond in different positions of an 18-carbon acyl chain were tested on AChR. Patch-clamp functional studies showed that only *cis*-6-18:1 and *cis*-9-18:1 reduce the duration of the AChR open state, thus suggesting an allosteric blocking mechanism. Fluorescence spectroscopy measurements demonstrated that all FFA locate in the AChR-lipid interphase, with *cis*-6-18:1 restricted to anular sites, while the rest of the FFA tested also occupy non-anular sites. Fluorescence polarization studies showed that *cis*-9-18:1 causes the highest membrane disorder of all FFA tested. It was determined that i) all *cis*-FFA generate AChR conformational changes at a transmembrane level, ii) only *cis*-9-18:1, *cis*-11-18:1 and *cis*-13-18:1 disturb AChR resting state and iii) *cis*-6-18:1 and *cis*-9-18:1 are the ones that cause the highest disturbance of the desensitized state. Thus, the position and isomerism of the torsion angle of unsaturated FFAs are probably a key factor in terms of AChR blockage, possibly by perturbing a transmembrane aminoacidic sequence involved in the allosteric changes necessary for ion channel gating.

In the plasma membrane, AChR is postulated to be located in lipid domains known as rafts. However, AChR shows no preference for L_o domains in model systems – composed of sphingomyelin (SM), cholesterol (Chol) and POPC (1:1:1) –, but a transmembrane segment (γ M4) that in closest contact with lipids does have

preference for them. This means that AChR distribution seems not to exclusively depend on its intrinsic properties but on signals external to the protein. In this Ph. D. thesis a possible differential AChR partitioning in L_o domains was studied in two model systems as a function of a) the presence of different pure SM species in the membrane and b) the existence of transbilayer asymmetry in the model system, by the addition of brain SM (bSM) in the external hemilayer. Both asymmetry and the presence of either 16:0-SM or 18:0-SM, in comparison with bSM or 24:1-SM, lead to an AChR preferential partitioning in L_o domains. AChR location in these domains depends not only on its properties but also on the characteristics of the membrane in which the ion channel is immersed.

Understanding lipid-AChR interaction is of great importance to determine treatments that can either improve or inhibit AChR function and, this, in turn, will help determining the treatment of diseases in which AChR is involved.

BIBLIOGRAFÍA

- Abràmoff M., Magalhães P., y Ram S. Image processing with imagej. *Biophotonics International*, 11(7): 36–42, 2004.
- Andersen N., Corradi J., Sine S. M., y Bouzat C. Stoichiometry for activation of neuronal $\alpha 7$ nicotinic receptors. *Proceedings of the National Academy of Sciences*, 110(51):20819–20824, 2013.
- Andersen O. S. Membrane proteins: Through thick and thin. *Nature Chemical Biology*, 9:667–668, 2013.
- Andreasen T. y McNamee M. Inhibition of ion permeability control properties of acetylcholine receptor from *Torpedo californica* by long-chain fatty acids. *Biochemistry*, 19(20):4719–4726, 1980.
- Andreeva I., Nirthanan S., Cohen J., y Pedersen S. Site specificity of agonist-induced opening and desensitization of the *Torpedo californica* nicotinic acetylcholine receptor. *Biochemistry*, 45(1):195–204, 2006.
- Antollini S., Soto M., de Romanelli I., Gutierrez-Merino C., Sotomayor P., y Barrantes F. Physical state of bulk and protein-associated lipid in nicotinic acetylcholine receptor-rich membrane studied by laurdan generalized polarization and fluorescence energy transfer. *Biophysical Journal*, 70(3):1275, 1996.
- Antollini S. S., y Barrantes F. J. Disclosure of discrete sites for phospholipid and sterols at the protein-lipid interface in native acetylcholine receptor-rich membrane. *Biochemistry*, 37(47):16653–16662, 1998.
- Antollini S. S., y Barrantes F. J. Unique effects of different fatty acid species on the physical properties of the torpedo acetylcholine receptor membrane. *Journal of Biological Chemistry*, 277(2):1249–1254, 2002.
- Antollini S. S., y Barrantes F. J. Laurdan Studies of Membrane Lipid-Acetylcholine Receptor Protein Interactions. Cap. 36. En: *Methods in Membrane Lipids* (A.M. Dopico, ed.). Humana Press Inc., Totowa, New Jersey, EE.UU., pp. 531–542, 2007.
- Antollini S., Wenz J., y Barrantes F. J. Cholesterol, fatty acids and nicotinic acetylcholine receptors. *Signal Transduction in Nervous Cells*, Research Signpost, Kerala, India, 3:39–61, 2009.
- Auerbach A. y Akk G. Desensitization of mouse nicotinic acetylcholine receptor channels a two-gate mechanism. *The Journal of General Physiology*, 112(2):181–197, 1998.
- Auerbach A. Gating of acetylcholine receptor channels: brownian motion across a broad transition state. *Proceedings of the National Academy of Sciences of the United States of America*, 102(5):1408–1412, 2005.
- Baenziger J., Morris M.-L., Darsaut T., y Ryan S. Effect of membrane lipid composition on the conformational equilibria of the nicotinic acetylcholine receptor. *Journal of Biological Chemistry*, 275(2): 777–784, 2000.

- Baenziger J. E., y da Costa C. J. B. Molecular mechanisms of acetylcholine receptor–lipid interactions: from model membranes to human biology. *Biophysical Reviews*, 5:1–9, 2013.
- Bagatolli L., Ipsen J., Simonsen A., y Mouritsen O. An outlook on organization of lipids in membranes: searching for a realistic connection with the organization of biological membranes. *Progress in Lipid Research*, 49(4):378–389, 2010.
- Bai J. y Pagano R. Measurement of spontaneous transfer and transbilayer movement of bodipy-labeled lipids in lipid vesicles. *Biochemistry*, 36(29):8840–8848, 1997.
- Baier C. J., Fantini J. y Barrantes F. J. (2011). Disclosure of cholesterol recognition motifs in transmembrane domains of the human nicotinic acetylcholine receptor. *Scientific Reports*, 1:0069, 2011.
- Barenholz Y., y Thompson T. Sphingomyelins in bilayers and biological membranes. *Biochimica et Biophysica Acta (BBA)-Biomembranes*, 604(2):129–158, 1980.
- Barrantes F. J. Endogenous chemical receptors: Some physical aspects. *Annual Review of Biophysics and Bioengineering* 8:287321, 1979.
- Barrantes F. J. Interactions between the acetylcholine receptor and the non-receptor, peripheral nu-peptide (mr 43000). *Neuroreceptors*, W. de Gruyter, Berlin, pp. 315–328, 1982.
- Barrantes F.J. (1989). The lipid environment of the nicotinic receptor in native and reconstituted membranes. En: Crit. Rev. Biochem. Molec. Biol. 24 (G. Fassman, ed.) (CRC Press, Boca Raton, Fl., EE.UU.), pp. 437489.
- Barrantes F. J., Editor. *The Nicotinic Acetylcholine Receptor: Current Views and Future Trends*. Springer Verlag, Berlin/Heidelberg and Landes Publishing Co, 1998.
- Barrantes F. J., Antollini S. S., Blanton M. P., y Prieto M. Topography of nicotinic acetylcholine receptor membrane-embedded domains. *Journal of Biological Chemistry*, 275(48), 37333–37339, 2000.
- Barrantes F. J. Modulation of nicotinic acetylcholine receptor function through the outer and middle rings of transmembrane domains. *Current Opinion in Drug Discovery & Development*, 6(5):620–632, 2003.
- Barrantes F. J. Structural basis for lipid modulation of nicotinic acetylcholine receptor function. *Brain Research Reviews*, 47(1):71–95, 2004
- Barrantes F. J. Cholesterol effects on nicotinic acetylcholine receptor. *Journal of Neurochemistry*, 103(s1):72–80, 2007.
- Barrantes F. J. Cholesterol effects on nicotinic acetylcholine receptor: cellular aspects. *Cholesterol Binding and Cholesterol Transport Proteins*, Springer Verlag, Berlin, pp. 467–487, 2010.
- Barrantes F. J. Regulation of the nicotinic acetylcholine receptor by cholesterol as a boundary lipid. *Cholesterol Regulation of Ion Channels and Receptors*, John Wiley & Sons, NY, pp. 181–204, 2012.
- Bermúdez V., Antollini S. S, Fernandez Nieves G. A., Aveldaño M. I, y Barrantes F. J. Partition profile of the nicotinic acetylcholine receptor in lipid domains upon reconstitution. *Journal of Lipid Research*, 51(9):2629–2641, 2010.
- Bermúdez V. Mecanismo de asociación del receptor de acetilcolina nicotínico a su microentorno lipídico. *Tesis Doctoral* bajo la dirección de los Dres. Barrantes F.J. y Aveldaño M.I., Universidad Nacional del Sur, 2011.
- Bernard C. Action du curare et de la nicotine sur le système nerveux et sur le système musculaire. *Comptes Rendus des Séances et Mémoires de la Société de Biologie*, 2:195, 1850.

- Bertrand D., Picard F., Le Hellard S., Weiland S., Favre I., Phillips H., Bertrand S., Berkovic S., Malafosse A., y Mulley J. How mutations in the nachrs can cause adnfle epilepsy. *Epilepsia*, 43(s5):112–122, 2002.
- Björkbom A., Róg T., Kankaanpää P., Lindroos D., Kaszuba K., Kurita M., Yamaguchi S., Yamamoto T., Jaikishan S., Paavolainen L., Päivärinne J., Nyholm T. K. M., Katsumura S., Vattulainen I., Slotte J. P. N- and O-methylation of sphingomyelin markedly affects its membrane properties and interactions with cholesterol. *Biochimica et Biophysica Acta – Biomembranes*, 1808(4):1179–1186, 2011.
- Blanton M., Xie Y., Dangott L., y Cohen J. The steroid promegestone is a noncompetitive antagonist of thetorpedo nicotinic acetylcholine receptor that interacts with the lipid-protein interfase. *Molecular Pharmacology*, 55(2):269–278, 1999.
- Bligh E. y Dyer W. A rapid method of total lipid extraction and purification. *Canadian Journal of Biochemistry and Physiology*, 37(8):911–917, 1959.
- Blount P. y Merlie J. Molecular basis of the two nonequivalent ligand binding sites of the muscle nicotinic acetylcholine receptor. *Neuron*, 3(3):349–357, 1989.
- Bonini, I.C., Antollini, S.S., Gutiérrez-Merino, C. y Barrantes, F.J. Sphingomyelin composition and physical asymmetries in native acetylcholine receptor-rich membranes. *European Biophysics Journal*, 31:417427, 2002.
- Borroni V., Baier C. J., Lang T., Bonini I., White M. M., Garbus I., y Barrantes F. J. Cholesterol depletion activates rapid internalization of submicron-sized acetylcholine receptor domains at the cell membrane. *Molecular Membrane Biology*, 24(1):1-15, 2007.
- Bouzat C. y Barrantes F. J. Hydrocortisone and 11-desoxycortisone modify acetylcholine receptor channel gating. *Neuroreport*, 4(2):143–146, 1993a.
- Bouzat C. y Barrantes F. J. Effects of long-chain fatty acids on the channel activity of the nicotinic acetylcholine receptor. *Receptors & Channels*, 1(3):251, 1993b.
- Bouzat C., Barrantes F. J., y Sine S. Nicotinic receptor fourth transmembrane domain hydrogen bonding by conserved threonine contributes to channel gating kinetics. *The Journal of General Physiology*, 115(5):663–672, 2000.
- Bouzat C., Lacorazza H., Jiménez Bonino M., y Barrantes F. J. Effect of chemical modification of extracellular histidyl residues on the channel properties of the nicotinic acetylcholine receptor. *Pflügers Archiv European Journal of Physiology*, 423(5):365–371, 1993.
- Bouzat C., Roccamo A. M., Garbus I., y Barrantes F. J. Mutations at lipid-exposed residues of the acetylcholine receptor affect its gating kinetics. *Molecular Pharmacology*, 54(1):146–153, 1998.
- Brannigan G., Hénin J., Law R., Eckenhoff R., y Klein M. Embedded cholesterol in the nicotinic acetylcholine receptor. *Proceedings of the National Academy of Sciences*, 105(38):14418–14423, 2008.
- Brody A. L., Olmstead R. E., London E. D., Farahi J., Meyer J.H., Grossman P., Lee G.S., Huang J., Hahn E.L., y Mandelkern M.A. Smoking-induced ventral striatum dopamine release. *American Journal of Psychiatry*, 161:1211-1218, 2004.
- Brown D. y London E. Functions of lipid rafts in biological membranes. *Annual Review of Cell and Developmental Biology*, 14(1):111–136, 1998.
- Brown D. y Rose J. Sorting of gpi-anchored proteins to glycolipid-enriched membrane subdomains during transport to the apical cell surface. *Cell*, 68(3):533–544, 1992.

- Buccafusco J. J., Beach J. W., y Terry Jr. A. V. Desensitization of nicotinic acetylcholine receptors as a strategy for drug development. *Journal of Pharmacology and Experimental Therapeutics*, 328(2):364-370, 2009.
- Campagna J., y Fallon J. Lipid rafts are involved in c95 (4, 8) agrin fragment-induced acetylcholine receptor clustering. *Neuroscience*, 138(1):123-132, 2006.
- Changeux J.-P., Meunier J.-C., y Huchet M. Studies on the cholinergic receptor protein of *Electrophorus electricus* I. An assay *in vitro* for the cholinergic receptor site and solubilization of the receptor protein from electric tissue. *Molecular Pharmacology* 7(5):538-553, 1971.
- Changeux J.-P. The nicotinic acetylcholine receptor: The founding father of the pentameric ligand-gated ion channel superfamily. *Journal of Biological Chemistry*, 287:40207-40215, 2012.
- Cheng H. y London E. Preparation and properties of asymmetric large unilamellar vesicles: interleaflet coupling in asymmetric vesicles is dependent on temperature but not curvature. *Biophysical Journal*, 100(11):2671-2678, 2011.
- Cheng H., Megha E., y London E. Preparation and properties of asymmetric vesicles that mimic cell membranes effect upon lipid raft formation and transmembrane helix orientation. *Journal of Biological Chemistry*, 284(10):6079-6092, 2009.
- Chong P.-G., Zhu W., y Venegas B. On the lateral structure of model membranes containing cholesterol. *Biochimica et Biophysica Acta (BBA)-Biomembranes*, 1788(1):2-11, 2009.
- Cooper E., Couturier S., y Ballivet M. Pentameric structure and subunit stoichiometry of a neuronal nicotinic acetylcholine receptor. *Nature*, 350:235-238, 1991.
- Corringer P.-J., Novère N. L., y Changeux J.-P. Nicotinic receptors at the amino acid level. *Annual Review of Pharmacology and Toxicology*, 40(1):431-458, 2000.
- Criado M., Eibl H., y Barrantes F. J. Effects of lipids on acetylcholine receptor. essential need of cholesterol for maintenance of agonist-induced state transitions in lipid vesicles. *Biochemistry*, 21(15):3622-3629, 1982.
- Criado M., Eibl H., y Barrantes F. J. Functional properties of the acetylcholine receptor incorporated in model lipid membranes. Differential effects of chain length and head group of phospholipids on receptor affinity states and receptor-mediated ion translocation. *Journal of Biological Chemistry*, 259(14):9188-9198, 1984.
- Crossthwaite A. J., Seebacher T., Masada N., Ciruela A., Dufraux K., Schultz J. E., y Cooper D. M. F. The cytosolic domains of Ca^{2+} -sensitive adenylyl cyclases dictate their targeting to plasma membrane lipid rafts. *Journal of Biological Chemistry*, 280(8):6380-6391, 2005.
- da Costa C. J. B., Ogrel A. A., McCardy E. A., Blanton M. P., y Baenziger J. E. Lipid-protein interactions at the nicotinic acetylcholine receptor. *Journal of Biological Chemistry*, 277(1):201-208, 2002.
- da Costa C. J. B., Wagg I., McKay M., y Baenziger J. E. Phosphatidic acid and phosphatidylserine have distinct structural and functional interactions with the nicotinic acetylcholine receptor. *Journal of Biological Chemistry*, 279(15):14967-14974, 2004.
- da Costa C. J. B., Medaglia S., Lavigne N., Wang S., Carswell C., y Baenziger J. E. Anionic lipids allosterically modulate multiple nicotinic acetylcholine receptor conformational equilibria. *Journal of Biological Chemistry*, 284(49):33841-33849, 2009.
- da Costa C. J. B. y Baenziger J. E. A lipid-dependent uncoupled conformation of the acetylcholine receptor. *Journal of Biological Chemistry*, 284(26):17819-17825, 2009.

- da Costa C. J. B., Dey L., Therien, J. P. D., y Baenziger, J. E. A distinct mechanism for activating uncoupled nicotinic acetylcholine receptors. *Nature Chemical Biology*, 9(11):701–707, 2013.
- Dai Z., Luo X., Xie H., y Peng H. B. The actin-driven movement and formation of acetylcholine receptor clusters. *The Journal of Cell Biology*, 150(6):1321–1334, 2000.
- Daleke D. L. Regulation of transbilayer plasma membrane phospholipid asymmetry. *Journal of Lipid Research*, 44(2):233–242, 2003.
- de Almeida R. F., Fedorov A., y Prieto M. Sphingomyelin/phosphatidylcholine/cholesterol phase diagram: Boundaries and composition of lipid rafts. *Biophysical Journal*, 85(4):2406–2416, 2003.
- de Almeida R. F., Loura L., Prieto M., Watts A., Fedorov A., y Barrantes F. J. Cholesterol modulates the organization of the γ m4 transmembrane domain of the muscle nicotinic acetylcholine receptor. *Biophysical Journal*, 86(4):2261–2272, 2004.
- Devaux P. y Seigneuret M. Specificity of lipid-protein interactions as determined by spectroscopic techniques. *Biochimica et Biophysica Acta (BBA)-Reviews on Biomembranes*, 822(1):63–125, 1985.
- Devaux P. F. Static and dynamic lipid asymmetry in cell membranes. *Biochemistry*, 30(5):1163–1173, 1991.
- Devaux P. F. y Morris R. Transmembrane asymmetry and lateral domains in biological membranes. *Traffic*, 5(4):241–246, 2004.
- Diaz-Rohrer B. B., Levental K. R., Simons K., y Levental I. Membrane raft association is a determinant of plasma membrane localization. *Proceedings of the National Academy of Science*, 111(23):8500–8505, 2014.
- Dietrich C., Bagatolli L., Volovyk Z., Thompson N., Levi M., Jacobson K., y Gratton E. Lipid rafts reconstituted in model membranes. *Biophysical Journal*, 80(3):1417–1428, 2001.
- Dilger J. P. y Liu Y. Desensitization of acetylcholine receptors in bc3h-1 cells. *Pflügers Archiv*, 420(5-6): 479–485, 1992.
- Dunn M. Determination of total protein concentration. *Protein Purification Methods*. Oxford IRL, 1989.
- Duque D., Li X.-j., Katsov K., y Schick M. Molecular theory of hydrophobic mismatch between lipids and peptides. *arXiv preprint cond-mat/0108237*, 2001.
- Edelstein S., Schaad O., Henry E., Bertrand D., y Changeux J.-P. A kinetic mechanism for nicotinic acetylcholine receptors based on multiple allosteric transitions. *Biological Cybernetics*, 75(5):361–379, 1996.
- Elenes S. y Auerbach A. Desensitization of diliganded mouse muscle nicotinic acetylcholine receptor channels. *The Journal of Physiology*, 541(2):367–383, 2002.
- Ellena J. F., Blazing M. A., y McNamee M. G. Lipid-protein interactions in reconstituted membranes containing acetylcholine receptor. *Biochemistry*, 22(24):5523–5535, 1983.
- Elliott J., Blanchard S., Wu W., Miller J., Strader C., Hartig P., Moore H., Racs J., y Raftery M. Purification of Torpedo californica post-synaptic membranes and fractionation of their constituent proteins. *Biochemical Journal*, 185(3):667, 1980.
- Ellman G. Tissue sulfhydryl groups. *Archives of Biochemistry and Biophysics*, 82:70–77, 1959.
- Engel A. G. y Sine S. M. Current understanding of congenital myasthenic syndromes. *Current Opinion in Pharmacology*, 5(3):308–321, 2005.

- Engel A. The neuromuscular junction. *Handbook of Clinical Neurology*, 91:103–148, 2008.
- Engelman D. M. Membranes are more mosaic than fluid. *Nature*, 438(7068):578–580, 2005.
- Fantini J., y Barrantes F. J. Sphingolipid/cholesterol regulation of neurotransmitter receptor conformation and function. *Biochimica et Biophysica Acta*, 1788:2345–2361, 2009.
- Fantini J., y Barrantes F. J. How cholesterol interacts with membrane proteins: an exploration of cholesterol-binding sites including CRAC, CARC, and tilted domains. *Frontiers in Physiology*, 4, 2013.
- Fadok V. A., Voelker D. R., Campbell P. A., Cohen J. J., Bratton D. L., y Henson P. M. Exposure of phosphatidylserine on the surface of apoptotic lymphocytes triggers specific recognition and removal by macrophages. *The Journal of Immunology*, 148(7):2207–2216, 1992.
- Fernández Nievas G., Barrantes F., y Antollini S. Conformation-sensitive steroid and fatty acid sites in the transmembrane domain of the nicotinic acetylcholine receptor. *Biochemistry*, 46(11):3503–3512, 2007.
- Fernández Nievas G., Barrantes F., y Antollini S. Modulation of nicotinic acetylcholine receptor conformational state by free fatty acids and steroids. *Journal of Biological Chemistry*, 283(31):21478–21486, 2008.
- Fong T. M. y McNamee M. G. Correlation between acetylcholine receptor function and structural properties of membranes. *Biochemistry*, 25(4):830–840, 1986.
- Fong T. M. y McNamee M. G. Stabilization of acetylcholine receptor secondary structure by cholesterol and negatively charged phospholipids in membranes. *Biochemistry*, 26(13):3871–3880, 1987.
- Forman S. A. Anesthetic sites and allosteric mechanisms of action on cys-loop ligand-gated ion channels. *Canadian Journal of Anesthesia/Journal Canadien d'Anesthésie*, 58(2):191–205, 2011.
- Förster T. *Intermolecular energy transfer and fluorescence*. National Research Council of Canada, 1955.
- Fritzsche K. J., Kim J., y Holland G. P. Probing lipid-cholesterol interactions in DOPC/eSM/Chol and DOPC/DPPE/Chol model lipid rafts with DSC and ¹³C solid-state NMR. *Biochimica et Biophysica Acta*, 1828:1889–1898, 2013.
- Froehner S. C. The submembrane machinery for nicotinic acetylcholine receptor clustering. *The Journal of Cell Biology*, 114(1):1–7, 1991.
- Fukudome T., Ohno K., Brengman J. M., y Engel A. G. Quinidine normalizes the open duration of slow-channel mutants of the acetylcholine receptor. *Neuroreport*, 9(8):1907–1911, 1998.
- Gandhavadi M., Allende D., Vidal A., Simon S., y McIntosh T. Structure, composition, and peptide binding properties of detergent soluble bilayers and detergent resistant rafts. *Biophysical Journal*, 82(3):1469–1482, 2002.
- Garbus I., Bouzat C., y Barrantes F. Steroids differentially inhibit the nicotinic acetylcholine receptor. *Neuroreport*, 12(2):227–231, 2001.
- Garvik O., Benediktson P., Simonsen A. C., Ipsen J. H., y Wüstner D. The fluorescent cholesterol analog dehydroergosterol induces liquid-ordered domains in model membranes. *Chemistry and Physics of Lipids*, 159(2):114–118, 2009.
- Gautam M., Noakes P. G., Moscoso L., Rupp F., Scheller R. H., Merlie J. P., y Sanes J. R. Defective neuromuscular synaptogenesis in agrin-deficient mutant mice. *Cell*, 85:525–535, 1996.

- Giniatullin R., Nistri A., y Yakel J. Desensitization of nicotinic ach receptors: shaping cholinergic signaling. *Trends in Neurosciences*, 28(7):371–378, 2005.
- Glass D. J., Bowen D. C., Stitt T. N., Radziejewski C., Bruno J., Ryan T. E., Gies D. R., Shah S., Mattsson K., Burden S. J., *et al.* Agrin acts via a musk receptor complex. *Cell*, 85(4):513–523, 1996.
- Goñi F.M. The basic structure and dynamics of cell membranes: An update of the Singer–Nicolson model. *Biochimica et Biophysica Acta*, 1838:1467–1476, 2014.
- Gri G., Molon B., Manes S., Pozzan T., y Viola A. The inner side of t cell lipid rafts. *Immunology Letters*, 94(3):247–252, 2004.
- Gutierrez-Merino C., de Romanelli I., Pietrasanta L., y Barrantes F. Preferential distribution of the fluorescent phospholipid probes nbd-phosphatidylcholine and rhodamine-phosphatidylethanolamine in the exofacial leaflet of acetylcholine receptor-rich membranes from *Torpedo marmorata*. *Biochemistry*, 34(14):4846–4855, 1995.
- Hall Z. W. y Sanes J. R. Synaptic structure and development: the neuromuscular junction. *Cell*, 72:99–121, 1993.
- Hamilton S., Pratt D., y Eaton D. Arrangement of the subunits of the nicotinic acetylcholine receptor of *Torpedo californica* as determined by α -neurotoxin crosslinking. *Biochemistry*, 24(9):2210–2219, 1985.
- Hammond A., Heberle F., Baumgart T., Holowka D., Baird B., y Feigenson G. Crosslinking a lipid raft component triggers liquid ordered-liquid disordered phase separation in model plasma membranes. *Proceedings of the National Academy of Sciences of the United States of America*, 102(18):6320–6325, 2005.
- Hamouda A. K., Sanghvi M., Sauls D., Machu T. K., y Blanton M. P. Assessing the lipid requirements of the *Torpedo californica* nicotinic acetylcholine receptor. *Biochemistry*, 45(13):4327–4337, 2006.
- Harder T. y Engelhardt K. R. Membrane domains in lymphocytes—from lipid rafts to protein scaffolds. *Traffic*, 5(4):265–275, 2004.
- Harper C. M., Fukudome T., y Engel A. G. Treatment of slow-channel congenital myasthenic syndrome with fluoxetine. *Neurology*, 60(10):1710–1713, 2003.
- Harris F., Best K., y Bell J. Use of laurdan fluorescence intensity and polarization to distinguish between changes in membrane fluidity and phospholipid order. *Biochimica et Biophysica Acta (BBA) - Biomembranes*, 1565(1):123–128, 2002.
- Hartig P. y Raftery M. Preparation of right-side-out, acetylcholine receptor enriched intact vesicles from *Torpedo californica* electroplaque membranes. *Biochemistry*, 18(7):1146–1150, 1979.
- Haydar S. N., y Dunlop J. Neuronal nicotinic acetylcholine receptors—targets for the development of drugs to treat cognitive impairment associated with schizophrenia and Alzheimer's disease. *Current Topics in Medicinal Chemistry*, 10(2):144–152, 2010.
- Hishikawa D., Shindou H., Kobayashi S., Nakanishi H., Taguchi R., y Shimizu T. Discovery of a lysophospholipid acyltransferase family essential for membrane asymmetry and diversity. *Proceedings of the National Academy of Sciences*, 105(8):2830–2835, 2008.
- Holloway P. A simple procedure for removal of triton x-100 from protein samples. *Analytical Biochemistry*, 53(1):304–308, 1973.
- Horváth L., Arias H., Hankovszky H., Hideg K., Barrantes F., y Marsh D. Association of spin-labeled local anesthetics at the hydrophobic surface of the acetylcholine receptor in native membranes from *Torpedo marmorata*. *Biochemistry*, 29(37):8707–8713, 1990.

- Huang, J., y Feigenson, G. W. A microscopic interaction model of maximum solubility of cholesterol in lipid bilayers. *Biophysical Journal*, 76(4):2142-2157, 1999.
- Hughes B. W., Kusner L. L., y Kaminski H. J. Molecular architecture of the neuromuscular junction. *Muscle & Nerve*, 33(4):445-461, 2006.
- Hurst R., Rollema H., y Bertrand D. Nicotinic acetylcholine receptors: From basic science to therapeutics. *Pharmacology & Therapeutics*, 137(1):22-54, 2013.
- Jaikishan S. y Slotte J. P. Effect of hydrophobic mismatch and interdigitation on sterol/sphingomyelin interaction in ternary bilayer membranes. *Biochimica et Biophysica Acta - Biomembranes*, 1808(7):1940-1945, 2011.
- Jain M. K. y White H. Long-range order in biomembranes. *Advances in Lipid Research*, 15:160, 1977.
- Jiménez-Garduño A. M., Mitkovski M., Alexopoulos I. K., Sánchez A., Stühmer W., Pardo L. A., y Ortega A. Kv10.1 channel plasma membrane discrete domain partitioning and its functional correlation in neurons. *Biochimica et Biophysica Acta - Biomembranes*, 1838(3):921-931, 2014.
- Jones O. T. y McNamee M. G. Annular and nonannular binding sites for cholesterol associated with the nicotinic acetylcholine receptor. *Biochemistry*, 27(7):2364-2374, 1988.
- Jones O. T., Eubanks J. H., Earnest J. P., y McNamee M. G. A minimum number of lipids are required to support the functional properties of the nicotinic acetylcholine receptor. *Biochemistry*, 27(10):3733-3742, 1988.
- Jonnala R. R. y Buccafusco J. Relationship between the increased cell surface $\alpha 7$ nicotinic receptor expression and neuroprotection induced by several nicotinic receptor agonists. *Journal of Neuroscience Research*, 66(4):565-572, 2001.
- Jungalwala F. B., Hayssen V., Pasquini J. M., y McCluer R. H. Separation of molecular species of sphingomyelin by reversed-phase high-performance liquid chromatography. *Journal of Lipid Research*, 20(5):579-587, 1979.
- Kaiser H.-J., Lingwood D., Levental I., Sampaio J. L., Kalvodova L., Rajendran L., y Simons K. Order of lipid phases in model and plasma membranes. *Proceedings of the National Academy of Sciences*, 106(39):16645-16650, 2009.
- Kaiser R. y London E. Location of diphenylhexatriene (dph) and its derivatives within membranes: comparison of different fluorescence quenching analyses of membrane depth. *Biochemistry*, 37(22):8180-8190, 1998.
- Karlin A. Emerging structure of the nicotinic acetylcholine receptors. *Nature Reviews Neuroscience*, 3(2):102-114, 2002.
- Karlin A., Holtzman E., Yodh N., Lobel P., Wall J., y Hainfeld J. The arrangement of the subunits of the acetylcholine receptor of torpedo californica. *Journal of Biological Chemistry*, 258(11):6678-6681, 1983.
- Karnovsky M. J., Kleinfeld A. M., Hoover R. L., y Klausner R. D. The concept of lipid domains in membranes. *The Journal of Cell Biology*, 94(1):1-6, 1982.
- Katz B. y Thesleff S. A study of the desensitization produced by acetylcholine at the motor end-plate. *The Journal of Physiology*, 138(1):63-80, 1957.
- Kellner R. R., Baier C. J., Willig K. I., Hell S. W., y Barrantes F. J. Nanoscale organization of nicotinic acetylcholine receptors revealed by stimulated emission depletion microscopy. *Neuroscience*, 144(1):135-143, 2007.
- Kiessling V., Crane J. M., y Tamm L. K. Transbilayer effects of raft-like lipid domains in asymmetric planar bilayers

- measured by single molecule tracking. *Biophysical Journal*, 91(9):3313–3326, 2006.
- Kirber M. T., Ordway R. W., Clapp L. H., Walsh Jr J. V., y Singer J. J. Both membrane stretch and fatty acids directly activate large conductance Ca^{2+} -activated K^+ channels in vascular smooth muscle cells. *FEBS Letters*, 297(1):24–28, 1992.
- Klausner R. D., Kleinfeld A. M., Hoover R. L., y Karnovsky M. J. Lipid domains in membranes. evidence derived from structural perturbations induced by free fatty acids and lifetime heterogeneity analysis. *Journal of Biological Chemistry*, 255(4):1286–1295, 1980.
- Kucerka N., Marquardt D., Harroun T. A., Nieh M. P., Wassall S. R., de Jong D. H., Schäfer L. V., Marrink S. J., y Katsaras J. Cholesterol in bilayers with PUFA chains: doping with DMPC or POPC results in sterol reorientation and membrane-domain formation. *Biochemistry*, 49(35):7485–7493, 2010.
- Laemmli U. Cleavage of structural proteins during the assembly of the head of bacteriophage t4. *Nature*, 227(5259):680–685, 1970.
- Lakowicz J. *Principles of Fluorescence Spectroscopy*, Volume 1. Springer, 2006.
- Langley J. On the contraction of muscle, chiefly in relation to the presence of “receptive” substances part i. *The Journal of Physiology*, 36(4-5):347–384, 1907.
- Lee A. How lipids affect the activities of integral membrane proteins. *Biochimica et Biophysica Acta - Biomembranes*, 1666(1):62–87, 2004.
- Lee Y., Li L., Lasalde J., Rojas L., McNamee M., Ortiz-Miranda S., y Pappone P. Mutations in the m4 domain of torpedo californica acetylcholine receptor dramatically alter ion channel function. *Biophysical Journal*, 66(3):646–653, 1994.
- Lentz B. y Burgess S. A dimerization model for the concentration dependent photophysical properties of diphenylhexatriene and its phospholipid derivatives. dphppc and dphppa. *Biophysical Journal*, 56(4):723–733, 1989.
- Lentz B., Wu J., Zheng L., y Prevratil J. The interfacial region of dipalmitoylphosphatidylcholine bilayers is perturbed by fusogenic amphipaths. *Biophysical Journal*, 71(6):3302–3310, 1996.
- Li L., Schuchard M., Palma A., Pradier L., y McNamee M. Functional role of the cysteine 451 thiol group in the m4 helix of the γ -subunit of Torpedo californica acetylcholine receptor. *Biochemistry*, 29(23):5428–5436, 1990.
- Lichtenberg D., Goñi F. M., y Heerklotz H. Detergent-resistant membranes should not be identified with membrane rafts. *Trends in Biochemical Sciences*, 30(8):430–436, 2005.
- Lin H., Hsu F. -C., Baumann B. H., Coulter D. A., y Lynch D. R. Cortical synaptic NMDA receptor deficits in $\alpha 7$ nicotinic acetylcholine receptor gene deletion models: Implications for neuropsychiatric diseases. *Neurobiology of Disease*, 63:129–140, 2014.
- Lin Q. y London E. Preparation of artificial plasma membrane mimicking vesicles with lipid asymmetry. *PloS one*, 9(1):e87903, 2014a.
- Lin Q. y London E. The influence of natural lipid asymmetry upon the conformation of a membrane- inserted protein (perfringolysin o). *Journal of Biological Chemistry*, 289:5467–5478, 2014b.
- Lindstrom J. Nicotinic acetylcholine receptors in health and disease. *Molecular Neurobiology*, 15(2):193–222, 1997.
- Lingwood D. y Simons K. Lipid rafts as a membrane-organizing principle. *Science*, 327(5961):46–50, 2010.

- Lowry O., Rosebrough N., Farr A., Randall R., *et al.*. Protein measurement with the folin phenol reagent. *Journal of Biological Chemistry*, 193(1):265–275, 1951.
- Lurtz M. y Pedersen S. Aminotriarylmethane dyes are high-affinity noncompetitive antagonists of the nicotinic acetylcholine receptor. *Molecular Pharmacology*, 55(1):159–167, 1999.
- Mantipragada S., Horvath L., Arias H., Schwarzmann G., Sandhoff K., Barrantes F., y Marsh D. Lipid- protein interactions and effect of local anesthetics in acetylcholine receptor-rich membranes from *Torpedo marmorata* electric organ. *Biochemistry*, 42(30):9167–9175, 2003.
- Marchand S., Devillers-Thiéry A., Pons S., Changeux J.-P., y Cartaud J. Rapsyn escorts the nicotinic acetylcholine receptor along the exocytic pathway via association with lipid rafts. *The Journal of Neuroscience*, 22(20):8891–8901, 2002.
- Marsh D. y Barrantes F. Immobilized lipid in acetylcholine receptor-rich membranes from *Torpedo marmorata*. *Proceedings of the National Academy of Sciences*, 75(9):4329–4333, 1978.
- Marsh D. y Horváth L. Structure, dynamics and composition of the lipid-protein interphase. Perspectives from spin-labelling. *Biochimica et Biophysica Acta (BBA)-Reviews on Biomembranes*, 1376(3):267–296, 1998.
- Marsh D., Watts A., y Barrantes F. Phospholipid chain immobilization and steriod rotational immobilization in acetylcholine receptor-rich membranes from *Torpedo marmorata*. *Biochimica et Biophysica Acta (BBA)-Biomembranes*, 645(1):97–101, 1981.
- Martinez-Seara H., Róg T., Karttunen M., Vattulainen I., y Reigada R. Cholesterol induces specific spatial and orientational order in cholesterol/phospholipid membranes. *PloS one*, 5(6):e11162, 2010.
- McIntosh T. J., Simon S. A., Needham D., y Huang C. H. Structure and cohesive properties of sphingomyelin/cholesterol bilayers. *Biochemistry*, 31(7):2012–2020, 1992.
- McIntosh T. J. Organization of skin stratum corneum extracellular lamellae: diffraction evidence for asymmetric distribution of cholesterol. *Biophysical Journal*, 85(3):1675–1681, 2003.
- McMahan U. J. The agrin hypothesis. *Cold Spring Harbor Symposia in Quantitative Biology*, 55:407–418, 1990.
- Milone M. y Engel A. G. Block of the endplate acetylcholine receptor channel by the sympathomimetic agents ephedrine, pseudoephedrine, and albuterol. *Brain Research*, 740(1):346–352, 1996.
- Minota S. y Watanabe S. Inhibitory effects of arachidonic acid on nicotinic transmission in bullfrog sympathetic neurons. *Journal of Neurophysiology*, 78(5):2396–2401, 1997.
- Mitra A., Bailey T., y Auerbach A. Structural dynamics of the m4 transmembrane segment during acetylcholine receptor gating. *Structure*, 12(10):1909–1918, 2004.
- Miyazawa A., Fujiyoshi Y., y Unwin N. Structure and gating mechanism of the acetylcholine receptor pore. *Nature*, 423(6943):949–955, 2003.
- Murakami M., y Kudo I. Phospholipase A₂. *Journal of Biochemistry*, 131:285–292, 2002.
- Narayanaswami V. y McNamee M. Protein-lipid interactions and *Torpedo californica* nicotinic acetylcholine receptor function. 2. Membrane fluidity and ligand-mediated alteration in the accessibility of γ subunit cysteine residues to cholesterol. *Biochemistry*, 32(46):12420–12427, 1993.
- Narayanaswami V., Kim J., y McNamee M. Protein-lipid interactions and *Torpedo californica* nicotinic acetylcholine receptor function. 1. Spatial disposition of cysteine

- residues in the γ subunit analyzed by fluorescence-quenching and energy-transfer measurements. *Biochemistry*, 32(46):12413–12419, 1993.
- Nicholson G. L. The fluid—mosaic model of membrane structure: Still relevant to understanding the structure, function and dynamics of biological membranes after more than 40 years. *Biochimica et Biophysica Acta*, 1838:1451–1466, 2014.
- Niemelä P., Hyvönen M. T., y Vattulainen I. Structure and dynamics of sphingomyelin bilayer: insight gained through systematic comparison to phosphatidylcholine. *Biophysical Journal*, 87(5):2976–2989, 2004.
- Niemelä P. S., Hyvönen M. T., y Vattulainen I. Influence of chain length and unsaturation on sphingomyelin bilayers. *Biophysical Journal*, 90(3):851–863, 2006.
- Ochoa E. L. M., Dalziel A. W., y McNamee M. G. Reconstitution of acetylcholine receptor function in lipid vesicles of defined composition. *Biochimica et Biophysica Acta (BBA) - Biomembranes*, 727(1): 151–162, 1983.
- Ochoa E. L. M., Chattopadhyay A., y McNamee M. G. Desensitization of the nicotinic acetylcholine receptor: Molecular mechanisms and effect of modulators. *Cellular and Molecular Neurobiology*, 9(2):141–178, 1989.
- Ordway R. W., Singer J. J., y Walsh Jr J. V. Direct regulation of ion channels by fatty acids. *Trends in Neurosciences*, 14(3):96–100, 1991.
- Ortells M. O. y Lunt G. G. Evolutionary history of the ligand-gated ion-channel superfamily of receptors. *Trends in Neurosciences*, 18(3):121–127, 1995.
- Ortells M. O., Barrantes G. E., Wood C., Lunt G. G., y Barrantes F. J. Molecular modelling of the nicotinic acetylcholine receptor transmembrane region in the open state. *Protein Engineering*, 10(5):511–517, 1997.
- Ortiz-Miranda S. I., Lasalde J. A., Pappone P. A., y McNamee M. G. Mutations in the m4 domain of the Torpedo californica nicotinic acetylcholine receptor alter channel opening and closing. *Journal of Membrane Biology*, 158(1):17–30, 1997.
- Parasassi T., De Stasio G., d'Ubaldo A., y Gratton E. Phase fluctuation in phospholipid membranes revealed by laurdan fluorescence. *Biophysical Journal*, 57(6):1179, 1990.
- Parasassi T., De Stasio G., Ravagnan G., Rusch R., y Gratton E. Quantitation of lipid phases in phospholipid vesicles by the generalized polarization of laurdan fluorescence. *Biophysical Journal*, 60(1):179, 1991.
- Parasassi T., Krasnowska E. K., Bagatolli L., y Gratton E. Laurdan and Prodan as polarity-sensitive fluorescent membrane probes. *Journal of Fluorescence*, 8(4):365–373, 1998.
- Peñalva D. A., Furland N. E., Lopez G. H., Avelano M. I., y Antollini S. S. Unique thermal behavior of sphingomyelin species with nonhydroxy and 2-hydroxy very-long-chain (c28-c32) polyunsaturated fatty acids. *Journal of Lipid Research*, 54:2225–2235, 2013.
- Perillo V. L., Fernández-Nievas G. A., Vallés A. S., Barrantes F. J., y Antollini S. S. The position of the double bond in monounsaturated free fatty acids is essential for the inhibition of the nicotinic acetylcholine receptor. *Biochimica et Biophysica Acta (BBA)-Biomembranes*, 1818(11):2511–2520, 2012.
- Picciotto M. R., Addy N. A., Mineur Y. S., y Brunzell D. H. It's not “either/or”: activation and desensitization of nicotinic acetylcholine receptors both contribute to behaviors related to nicotine addiction and mood. *Progress in Neurobiology*. 84(4): 329–342, 2008.
- Piguet J., Schreiter C., Segura J.-M., Vogel H., y Hovius R. Acetylcholine receptor

- organization in membrane domains in muscle cells: evidence for rapsyn-independent and rapsyn-dependent mechanisms. *Journal of Biological Chemistry*, 286:363-369, 2011.
- Pike L. J. Rafts defined: a report on the keystone symposium on lipid rafts and cell function. *Journal of Lipid Research*, 47(7):1597-1598, 2006.
- Pike L. J. The challenge of lipid rafts. *Journal of Lipid Research*, 50(Supplement):S323-S328, 2009.
- Pokorny A., Yandek L. E., Elegbede A. I., Hinderliter A., y Almeida P. F. F. Temperature and composition dependence of the interaction of δ -lysine with ternary mixtures of Sphingomyelin/Cholesterol/POPC. *Biophysical Journal*, 91(6):2184-2197, 2006.
- Purohit P., Gupta S., Jadey S., y Auerbach A. Functional anatomy of an allosteric protein. *Nature Communications*, 4:1-12, 2013.
- Quinn P. J. *Membrane dynamics and domains*, volume 37. Springer, Berlin, 2004.
- Rafferty M. A., Vandlen R. L., Reed K. L., y Lee T. Characterization of Torpedo californica acetylcholine receptor: its subunit composition and ligand-binding properties. En: *Cold Spring Harbor Symposia on Quantitative Biology*. Cold Spring Harbor Laboratory Press, 40, pp. 193-202, 1976.
- Raggers R. J., Pomorski T., Holthuis J., Kälin N., y van Meer G. Lipid traffic: the abc of transbilayer movement. *Traffic*, 1(3):226-234, 2000.
- Rajendran L. y Simons K. Lipid rafts and membrane dynamics. *Journal of Cell Science*, 118(6):1099-1102, 2005.
- Ramarao M. K. y Cohen J. B. Mechanism of nicotinic acetylcholine receptor cluster formation by rapsyn. *Proceedings of the National Academy of Sciences*, 95(7):4007-4012, 1998.
- Reiss K., Cornelsen I., Husmann M., Gimpl G., y Bhakdi S. Unsaturated fatty acids drive ADAM-dependent cell adhesion, proliferation and migration by modulating membrane fluidity. *Journal of Biological Chemistry*, 286:26931-26942, 2011.
- Rothman J. E. y Lenard J. Membrane asymmetry. *Science*, 195(4280):743-753, 1977.
- Rotstein N.P., Arias H.R., Barrantes F.J. y Avelaño M.I. Composition of lipids in elasmobranch electric organ and acetylcholine receptor membranes. *Journal of Neurochemistry*, 49:1333-1340, 1987.
- Rouser G., Fleischer S., y Yamamoto A. Two dimensional thin layer chromatographic separation of polar lipids and determination of phospholipids by phosphorus analysis of spots. *Lipids*, 5(5):494-496, 1970.
- Ryan S., Demers C., Chew J., y Baenziger J. Structural effects of neutral and anionic lipids on the nicotinic acetylcholine receptor an infrared difference spectroscopy study. *Journal of Biological Chemistry*, 271(40):24590-24597, 1996.
- Sakmann B. y Neher E. Single channel currents recorded from membrane of denervated frog muscle fibers. *Nature*, 260(799-802):7, 1976.
- Sanes J. R. y Lichtman J. W. Induction, assembly, maturation and maintenance of a postsynaptic apparatus. *Nature Reviews Neuroscience*, 2(11):791-805, 2001.
- Sanes J. y Lichtman J. Development of the vertebrate neuromuscular junction. *Annual Review of Neuroscience*, 22(1):389-442, 1999.
- Schara U., Della Marina A., y Abicht A. Congenital myasthenic syndromes: current diagnostic and therapeutic approaches. *Neuropediatrics*, 43(04):184-193, 2012.
- Scheiffele P., Roth M. G., y Simons K. Interaction of influenza virus haemagglutinin with sphingolipid-

- cholesterol membrane domains via its transmembrane domain. *The EMBO Journal*, 16(18):5501–5508, 1997.
- Schofield P. R., Darlison M. G., Fujita N., Burt D. R., Stephenson F. A., Rodriguez H., Rhee L. M., Ramachandran J., Reale V., y Glencorse T. A. Sequence and functional expression of the GABA A receptor shows a ligand-gated receptor super-family. *Nature*, 328:221–227, 1987.
- Schroeder F., Nemezc G., Gibson Wood W., Joiner C., Morrot G., Ayrault-Jarrier M., y Devaux P. F. Transmembrane distribution of sterol in the human erythrocyte. *Biochimica et Biophysica Acta (BBA)-Biomembranes*, 1066(2):183–192, 1991.
- Schroit A. J. y Zwaal R. F. A. Transbilayer movement of phospholipids in red cell y platelet membranes. *Biochimica et Biophysica Acta (BBA)-Reviews on Biomembranes*, 1071(3):313–329, 1991.
- Schwartz A. What's next for Alzheimer treatment? *Annals of Neurology*, 73(4):A7–A9, 2013.
- Shahidullah K., Krishnakumar S. S., y London E. The effect of hydrophilic substitutions y anionic lipids upon the transverse positioning of the transmembrane helix of the erbb2 *neu* protein incorporated into model membrane vesicles. *Journal of Molecular Biology*, 396(1):209–220, 2010.
- Sharpe H. J., Stevens T. J., y Munro S. A comprehensive comparison of transmembrane domains reveals organelle-specific properties. *Cell*, 142(1):158–169, 2010.
- Shen X.-M., Deymeer F., Sine S. M., y Engel A. G. Slow-channel mutation in acetylcholine receptor *am4* domain and its efficient knockdown. *Annals of Neurology*, 60(1):128–136, 2006.
- Shinitzky M. y Yuli I. Lipid fluidity at the submacroscopic level: determination by fluorescence polarization. *Chemistry and Physics of Lipids*, 30(2):261–282, 1982.
- Shipley, G. G. Recent X-ray diffraction studies of biological membranes and membrane components. *Biological Membranes*, 2:1–89, 1973.
- Silvius J. R. Role of cholesterol in lipid raft formation: lessons from lipid model systems. *Biochimica et Biophysica Acta (BBA) - Biomembranes*, 1610(2):174–183, 2003.
- Simons K. y Ikonen E. Functional rafts in cell membranes. *Nature*, 387(6633):569–572, 1997.
- Simons K. y Toomre D. Lipid rafts and signal transduction. *Nature Reviews Molecular Cell Biology*, 1(1):31–39, 2000.
- Simons K. y Vaz W. Model systems, lipid rafts, and cell membranes 1. *Annual Review of Biophysics and Biomolecular Structure*, 33:269–295, 2004.
- Sine S. M. y Engel A. G. Recent advances in cys-loop receptor structure and function. *Nature*, 440 (7083):448–455, 2006.
- Sine S. End-plate acetylcholine receptor: structure, mechanism, pharmacology, and disease. *Physiological Reviews*, 92(3):1189–1234, 2012.
- Sine S. y Steinbach J. Activation of acetylcholine receptors on clonal mammalian bc3h-1 cells by low concentrations of agonist. *The Journal of Physiology*, 373(1):129–162, 1986.
- Singer G., S.J. and Nicolson. The fluid mosaic model of the structure of cell membranes. *Science*, 175(23):720–731, 1972.
- Smotrys J. E. y Linder M. E. Palmitoylation of intracellular signaling proteins: regulation and function. *Annual Review of Biochemistry*, 73(1):559–587, 2004.
- Sousa C., Nunes C., Lúcio M., Ferreira H., Lima J., Tavares J., Cordeiro-da Silva A., y Reis S. Effect of nonsteroidal anti-

- inflammatory drugs on the cellular membrane fluidity. *Journal of Pharmaceutical Sciences*, 97(8):3195–3206, 2007.
- Steck T., Ye J., y Lange Y. Probing red cell membrane cholesterol movement with cyclodextrin. *Biophysical Journal*, 83(4):2118–2125, 2002.
- Stern O., y Volmer M. On the quenching-time of fluorescence. *Physik Zeitschr*, 20:183–188, 1919.
- Stetzkowski-Marden F., Recouvreur M., Camus G., Cartaud A., Marchand S., y Cartaud J. Rafts are required for acetylcholine receptor clustering. *Journal of Molecular Neuroscience*, 30(1):37–38, 2006.
- Subczynski W. y Kusumi A. Dynamics of raft molecules in the cell and artificial membranes: approaches by pulse epr spin labeling and single molecule optical microscopy. *Biochimica et Biophysica Acta (BBA)-Biomembranes*, 1610(2):231–243, 2003.
- Sunshine C. y McNamee M. Lipid modulation of nicotinic acetylcholine receptor function: the role of membrane lipid composition and fluidity. *Biochimica et Biophysica Acta (BBA)-Biomembranes*, 1191(1):59–64, 1994.
- Tamamizu S., Lee Y., Hung B., McNamee M., y Lasalde-Dominicci J. Alteration in ion channel function of mouse nicotinic acetylcholine receptor by mutations in the m4 transmembrane domain. *Journal of Membrane Biology*, 170(2):157–164, 1999.
- Tamamizu S., Guzmán G. R., Santiago J., Rojas L. V., McNamee M. G., y Lasalde-Dominicci J. A. Functional effects of periodic tryptophan substitutions in the α m4 transmembrane domain of the Torpedo californica nicotinic acetylcholine receptor. *Biochemistry*, 39(16):4666–4673, 2000.
- Thanvi B. y Lo T. Update on myasthenia gravis. *Postgraduate Medical Journal*, 80(950):690–700, 2004.
- Thompson A. J., Lester H. A. y Lummis S. C. R. The structural basis of function in Cys-loop receptors. *Quarterly Reviews of Biophysics*, 43:449–499, 2010.
- Tobimatsu T., Fujita Y., Fukuda K., Tanaka K.-I., Mori Y., Konno T., Mishina M., y Numa S. Effects of substitution of putative transmembrane segments on nicotinic acetylcholine receptor function. *FEBS letters*, 222(1):56–62, 1987.
- Tuba Z., Maho S., y Sylvester V. Synthesis and structure-activity relationships of neuromuscular blocking agents. *Current Medicinal Chemistry*, 9(16):1507–1536, 2002.
- Tulenko T., Chen M., Mason P., y Mason R. Physical effects of cholesterol on arterial smooth muscle membranes: evidence of immiscible cholesterol domains and alterations in bilayer width during atherogenesis. *Journal of Lipid Research*, 39(5):947–956, 1998.
- Unwin N. Acetylcholine receptor channel imaged in the open state. *Nature*, 373(6509):37–43, 1995.
- Unwin N. Refined structure of the nicotinic acetylcholine receptor at 4Å resolution. *Journal of Molecular Biology*, 346(4):967–989, 2005.
- Unwin N., Miyazawa A., Li J., y Fujiyoshi Y. Activation of the nicotinic acetylcholine receptor involves a switch in conformation of the α subunits. *Journal of Molecular Biology*, 319(5):1165–1176, 2002.
- Unwin N., y Fujiyoshi Y. Gating movement of acetylcholine receptor caught by plunge-freezing. *Journal of Molecular Biology*, 422(5):617–634., 2012.
- Unwin N. Nicotinic acetylcholine receptor and the structural basis of neuromuscular transmission: insights from Torpedo postsynaptic membranes. *Quarterly Reviews of Biophysics*, 46(4):283–322, 2013.
- Valcarcel C., Dalla Serra M., Potrich C., Bernhart I., Tejuca M., Martinez D., Pazos

- F., Lanio M., y Menestrina G. Effects of lipid composition on membrane permeabilization by sticholysin i and ii, two cytolysins of the sea anemone *stichodactyla helianthus*. *Biophysical Journal*, 80(6):2761–2774, 2001.
- Varma R., y Mayor S. GPI-anchored proteins are organized in submicron domains at the cell surface. *Nature*, 394(6695):798-801, 1998.
- Wallace B. Agrin-induced specializations contain cytoplasmic, membrane, and extracellular matrix-associated components of the postsynaptic apparatus. *The Journal of Neuroscience*, 9(4):1294–1302, 1989.
- Wassall S. y Stillwell W. Polyunsaturated fatty acid–cholesterol interactions: Domain formation in membranes. *Biochimica et Biophysica Acta (BBA)-Biomembranes*, 1788(1):24–32, 2009.
- Willmann R., Pun S., Stallmach L., Sadasivam G., Santos A., Caroni P., y Fuhrer C. Cholesterol and lipid microdomains stabilize the postsynapse at the neuromuscular junction. *The EMBO Journal*, 25(17):4050–4060, 2006.
- Wood W. G., Igbavboa U., Müller W. E., y Eckert G. P. Cholesterol asymmetry in synaptic plasma membranes. *Journal of Neurochemistry*, 116(5):684–689, 2011.
- Wu S. y McConnell H. Phase separations in phospholipid membranes. *Biochemistry*, 14(4):847–854, 1975.
- Xu Y., Barrantes F. J., Luo X., Chen K., Shen J., y Jiang H. Conformational dynamics of the nicotinic acetylcholine receptor channel: a 35-ns molecular dynamics simulation study. *Journal of the American Chemical Society*, 127(4):1291–1299, 2005.
- Yamabhai M. y Anderson R. G. W. Second cysteine-rich region of epidermal growth factor receptor contains targeting information for caveolae/rafts. *Journal of Biological Chemistry*, 277(28):24843–24846, 2002.
- Yu W., So P., French T., y Gratton E. Fluorescence generalized polarization of cell membranes: a two-photon scanning microscopy approach. *Biophysical Journal*, 70(2):626–636, 1996.
- Zacharias D. A., Violin J. D., Newton A. C., y Tsien R. Y. Partitioning of lipid-modified monomeric GFPs into membrane microdomains of live cells. *Science*, 296(5569):913–916, 2002.
- Zanello L. P., Aztiria E., Antollini S., y Barrantes, F. J. Nicotinic acetylcholine receptor channels are influenced by the physical state of their membrane environment. *Biophysical Journal*, 70(5), 2155-2164, 1996.
- Zheng W. y Auerbach A. Decrypting the sequence of structural events during the gating transition of pentameric ligand-gated ion channels based on an interpolated elastic network model. *PLoS Computational Biology*, 7(1):e1001046, 2011.
- Zhu D., Xiong W. C., y Mei L. Lipid rafts serve as a signaling platform for nicotinic acetylcholine receptor clustering. *Journal of Neuroscience*, 26(18):4841–4851, 2006.
- Zolese G., Gratton E., y Curatola G. Phosphatidic acid affects structural organization of phosphatidylcholine liposomes. A study of 1, 6-diphenyl-1, 3, 5-hexatriene (DPH) and 1-(4-trimethylammonium-phenyl)-6-phenyl, 1, 3, 5-hexatriene (TMA-DPH) fluorescence decay using distributional analysis. *Chemistry and Physics of Lipids*, 55(1):29–39, 1990.
- Zong Y., Zhang B., Gu S., Lee K., Zhou J., Yao G., Figueiredo D., Perry K., Mei L., y Jin R. Structural basis of agrin–LRP4–MuSK signaling. *Genes & Development*, 26: 247-258, 2012.